



New diterpenoids from *Caesalpinia* species and their cytotoxic activity[☆]

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ABSTRACT

Chemical investigation on *Caesalpinia crista* afforded two new diterpenoids, 6 β -cinnamoyloxy-7 β -acetoxycoumarin-5 α -ol and 6 β ,7 β -dibenzoyloxyvouacapen-5 α -ol and on *Caesalpinia pulcherrima* another new diterpenoid, 12-demethyl neocaesalpin F along with several known constituents. The structures of the new compounds were settled from their 1D and 2D NMR spectral data. The cytotoxicity of these compounds was measured on two different cancer cell lines.

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The species belong to *Caesalpinia* (Leguminosae) are evergreen and deciduous trees and shrubs grown as ornamental plants in tropical and subtropical countries. These plants are known for their medicinal properties, such as *Caesalpinia crista* is used¹ as tonic for the treatment of rheumatism and backache while *Caesalpinia pulcherrima* is applied² as abortifacient and emmenagogue. The metabolites of these two species are mainly diterpenoids^{3–7} flavonoids^{8,9} and peltogynoids.⁸ Some of the constituents are known to possess antitumor, antimicrobial and antimaterial properties.^{4–6} Here, we report the isolation, structure elucidation and cytotoxicity of two new diterpenoids, **1** and **2** from *C. crista* and of another new diterpenoid, **3** from *C. pulcherrima*.

Compound **1** was isolated as a white solid.¹⁰ Its molecular formula was decided to be C₃₁H₃₈O₆ from its HREIMS (*m/z* 529.2578 for C₃₁H₃₈O₆Na). The IR spectrum showed the presence of carbonyl and hydroxyl groups as well as unsaturation in the molecule. The structure of the compound was established from its ¹H and ¹³C NMR spectral data (Table 1) which indicated it to be a cassane diterpenoid.^{7,8} The proton and carbon signals were clearly assigned with the help of 2D NMR (¹H–¹H COSY, NOESY, HSQC and HMBC) and DEPT experiments. The spectral data suggested that the structure of **1** was related to 6 β -cinnamoyloxy-7 β -hydroxyvouacapen-

5 α -ol (**4**)³ (Fig. 1), previously reported from *C. pulcherrima*. However, the former contained an acetoxy group instead of the hydroxyl group present at C-7 in **4**. The acetoxy group was reasonably placed at C-7 in **1** as H-7 appeared downfield (δ 5.52, dd, *J* = 10.0, 4.0 Hz) and was related to H-6 (δ 5.72, d, *J* = 4.0 Hz) as indicated by coupling constant. Both the compounds **1** and **4** contained a *trans*-cinnamoyloxy moiety at C-6. The HMBC experiment (Fig. 2) showed that H-6 was correlated to C-1' (δ 166.1) which in turn was related to C-3' (δ 145.4). Similarly, H-7 was related to C-1'' (δ 170.5). The coupling constant (4.0 Hz) between H-6 and H-7 and the NOESY correlation clearly suggested their β -configuration. The ester groups of **1** were hydrolyzed with methanolic KOH to afford a known compound, 6 β -hydroxyisovouacapenol C (**5**)¹¹ which was characterized by comparison of its physical (mp and optical rotation) and spectral (IR, ¹H NMR and MS) data with those reported earlier. Thus the compound **1** was characterized as 6 β -cinnamoyloxy-7 β -acetoxycoumarin-5 α -ol.

Compound **2** was obtained as a white solid.¹⁰ Its molecular formula was decided to be C₃₄H₃₈O₆ from its HREIMS (*m/z* 565.2555 for C₃₄H₃₈O₆Na). Its IR absorption peaks as well as ¹H and ¹³C NMR data (Table 1) suggested that its structure was related to that of **1** (Fig. 1). However, the ester moieties at C-6 and C-7 were different. In **2** two dibenzoyloxy groups were present at these two positions. The HMBC spectrum revealed that H-6 (δ 6.03, d, *J* = 4.0 Hz) was related to the carbonyl group (δ 166.1) of a benzoyloxy moiety while H-7 (δ 5.85, dd, *J* = 10.0, 4.0 Hz) to another

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Table 1
NMR spectral data of compound **1**, **2** and **3**^a

Position	Compound 1			Compound 2			Compound 3		
	¹ H NMR	Multiplicity (<i>J</i> in Hz)	¹³ C NMR	¹ H NMR	Multiplicity (<i>J</i> in Hz)	¹³ C NMR	¹ H NMR	Multiplicity (<i>J</i> in Hz)	¹³ C NMR
1	1.60(a)1.44(b)	M	35.0	1.70(a) 1.52(b)	m	35.0	1.60(a) 1.53(b)	m	34.7
2	1.70(a) 1.52(b)	M	18.1	1.74(a) 1.55(b)	m	18.2	1.70(a) 1.54(b)	m	18.1
3	1.69(a) 1.16(b)	M	37.7	1.76(a) 1.16(b)	m	37.6	1.67(a) 1.15(b)	m	37.6
4	—	—	41.2	—	—	39.3	—	—	39.2
5	—	—	77.3	—	—	77.3	—	—	77.97
6	5.72	d (4.0)	70.2	6.03	d (4.0)	70.9	5.78	d (4.0)	73.9
7	5.52	dd (10.0, 4.0)	71.6	5.85	dd (10.0, 4.0)	72.3	4.48	dd (10.0, 4.0)	68.0
8	2.26	M	35.4	2.66	m	37.2	1.89	m	41.9
9	2.54	M	37.1	2.46	m	35.8	2.49	m	36.9
10	—	—	39.2	—	—	41.2	—	—	40.8
11	2.57	M	21.7	2.63	m	21.8	2.26(a) 1.44(b)	m	37.7
12	—	—	149.2	—	—	149.8	—	—	106.1
13	—	—	121.6	—	—	121.9	—	—	171.5
14	2.83	M	27.4	2.87	m	27.5	3.37	m	32.1
15	6.19	d (2.0)	109.5	6.15	d (2.0)	109.5	5.70	s	113.6
16	7.21	d (2.0)	140.5	7.24	d (2.0)	140.5	—	—	173.3
17	0.98	d (7.0)	17.1	1.02	d (7.0)	17.1	1.26	d (7.0)	12.2
18	1.47	S	27.7	1.61	s	17.6	1.42	s	17.5
19	1.18	S	25.4	1.18	s	25.4	1.07	s	27.8
20	1.02	S	17.6	1.10	s	27.7	1.13	s	25.4
1'	—	—	166.1	—	—	166.1	—	—	167.5
2'	6.43	d (16.0)	117.9	—	—	129.7	—	—	129.9
3'	7.70	d (16.0)	145.4	7.80	dd (8.0, 2.0)	129.2	8.02	dd (8.0, 1.5)	129.6
4'	—	—	134.1	7.42	t (8.0)	128.1	7.45	t (8.0)	128.6
5'	7.58–7.50	M	128.9	7.46	br t (8.0)	132.8	7.59	t (8.0)	133.4
6'	7.43–7.37	M	128.2	7.42	t (8.0)	128.1	7.45	t (8.0)	128.6
7'	7.43–7.37	M	130.5	7.80	dd (8.0, 2.0)	129.2	8.02	dd (8.0, 1.5)	129.6
8'	7.43–7.37	M	128.2	—	—	—	—	—	—
9'	7.58–7.50	M	128.9	—	—	—	—	—	—
1''	—	—	170.5	—	—	171.2	—	—	—
2''	1.98	S	20.9	—	—	130.2	—	—	—
3''	—	—	—	7.89	dd (8.0, 2.0)	129.6	—	—	—
4''	—	—	—	7.49	t (8.0)	128.5	—	—	—
5''	—	—	—	7.57	brt (8.0)	133.8	—	—	—
6''	—	—	—	7.49	t (8.0)	128.5	—	—	—
7''	—	—	—	7.89	dd (8.0, 2.0)	129.6	—	—	—
OH-2	1.82	br s	—	1.98	br s	—	2.16	br s	—
OH-12	—	—	—	—	—	—	2.16	br s	—

^a The spectra were run in CDCl₃ at 600 MHz (¹H NMR) and 150 MHz (¹³C NMR).

carbonyl group of the second benzoyloxy moiety (δ 171.2). The NOESY experiment decided the β -configuration of both H-6 and H-7. The hydrolysis of the ester groups of **2** with methanolic KOH afforded the same known compound, 6 β -hydroxyisovouacapenol C (**5**)¹¹ as was obtained by hydrolysis of **1**. Thus the structure of **2** was settled as 6 β ,7 β -dibenzoyloxyvouacapen-5 α -ol.

Compound **3** was isolated as a white solid.¹⁰ Its molecular formula was established as C₂₇H₃₄O₇ from its HREIMS (*m/z* 471.2395 for C₂₇H₃₅O₇). The IR spectrum showed the presence of hydroxyl and carbonyl groups and aromatic residue. The ¹H and ¹³C NMR spectral data (Table 1) suggested the compound to be an α,β -butenolide cassane diterpenoid while the earlier two compounds **1** and **2** were furano cassane diterpenoid. The spectral data revealed that the compound **3** was structurally related to neocaesalpin F (**6**) (Fig. 3) previously reported¹¹ from the same species. However, the former contained no methoxy group present at C-12 in the latter. Instead, a hydroxyl group was present in **3** at the same position. In the ¹³C NMR spectrum C-12 appeared at δ 106.1 and in the HMBC spectrum H-15 (δ 5.70, s) showed correlation with this carbon. In the NOESY experiment (Fig. 4) Me-17 (δ 1.26, d, *J* = 7.0 Hz) was correlated to H-7 (δ 4.48, dd, *J* = 10.0,

4.0 Hz) and H-9 (δ 2.49, td, *J* = 10.0, 2.0 Hz) which indicated that the C-ring is in the chair form and consequently HO-12 is α -oriented.¹² Me-17 also showed NOESY correlation with HO-12 (δ 2.16, br s). Thus the structure of **3** was deduced to be 12-demethyl neocaesalpin F.

The known homoisoflavanoids, 7-O-methyl bonducellin¹³ sappanone A¹⁴ and 4'-O-methyl sappanone A¹⁵ were also isolated from *C. crista* and bonducellin⁹ and isobonducellin⁹ from *C. pulcherrima*. In addition to these compounds the latter species afforded the simple flavanoids, 5,7-dimethoxy flavanone 5,7-dimethoxy-3',4'-methylenedioxy flavanone and 2'-hydroxy-2,3,4',6'-tetramethoxy chalcone.⁹

Compounds **1–3** were tested for in vitro cytotoxic activity against two cancer lines, HL-60 (Human promyelocytic leukemia) and HeLa (Human cervical carcinoma) using the MTT assay according to the protocol of Mossman.¹⁶ Camptothecin and etoposide were considered as the positive control. All the three compounds (**1–3**) showed significant decrease in cell viability in the test cell lines in a concentration dependent manner. IC₅₀ value was measured with each cell line after four individual observations (Table 2). The cytotoxic activity of the test compounds was found to be

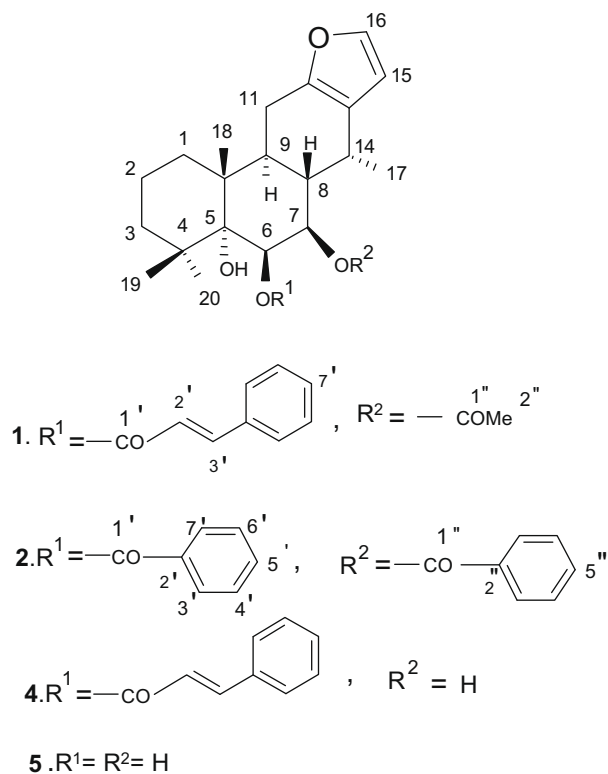


Figure 1.

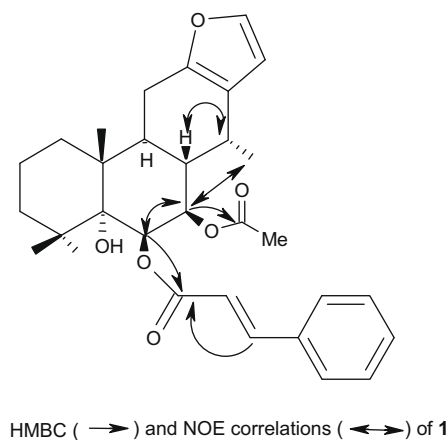


Figure 2.

Table 2
Cytotoxic activity of compounds 1–3 on two different cancer cell lines

Sample	IC ₅₀ ^a (μM)	
	HL-60	HeLa
Compound 1	17.37 ± 0.88	33.41 ± 0.75
Compound 2	19.77 ± 1.54	33.90 ± 0.66
Compound 3	44.89 ± 2.58	52.69 ± 2.48
Camptothecin	0.59 ± 0.04	0.94 ± 0.07
Etoposide	1.81 ± 0.20	8.93 ± 0.69

^a The results represent the mean ± standard error of four individual observations.

in the order of camptothecin > etoposide > 1 > 2 > 3 on both HL-60 and HeLa cells.

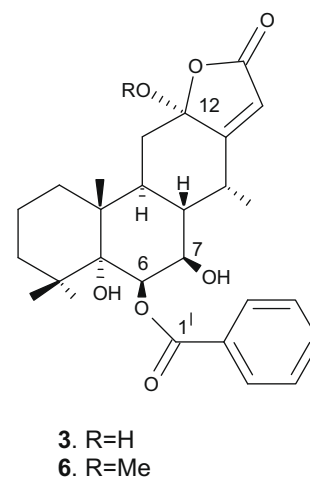
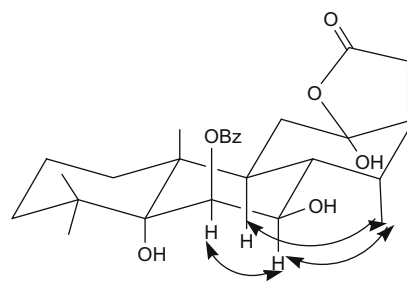


Figure 3.



conformation and NOE correlations (↔) of 3

Figure 4.

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References and notes

- Eisai, P. T. Indonesia Medicinal Herb Index in Indonesia, 1986, p 140.
- Quisimbing, E. *Medicinal Plants of the Philippines*; Bureau of Printing: Manila, 1951.
- McPherson, D. D.; Che, T. T.; Cordell, G. A.; Soejarto, D. D.; Pezzuto, J. M.; Fong, H. H. S. *Phytochemistry* **1986**, 25, 167.
- Che, C. T.; McPherson, D. D.; Cordell, G. A.; Fong, H. H. S. *J. Nat. Prod.* **1986**, 49, 561.
- Patel, A. D.; Freyer, A. J.; Webl, R. L.; Zuber, G.; Reichwein, R.; Bean, M. F.; Faucett, L.; Johnson, R. K. *Tetrahedron* **1977**, 53, 1583.
- Ragasa, C. Y.; Hofilena, J. G.; Rideout, J. A. *J. Nat. Prod.* **2002**, 65, 1107.
- Kalauni, S. K.; Awale, S.; Tezuka, Y.; Banskola, A. H.; Luin, T. Z.; Kadota, S. *Chem. Pharm. Bull.* **2005**, 53, 214.
- McPherson, D. D.; Cordell, G. A.; Soejarto, D. D.; Pezzuto, J. M.; Fong, H. H. S. *Phytochemistry* **1983**, 27, 2835.
- Srinivas, K. V. N. S.; Rao, Y. K.; Mahender, I.; Das, B.; Krishna, K. V. S. R.; Kishore, K. H.; Murthy, U. S. N. *Phytochemistry* **2003**, 63, 789.
- Isolation of constituents of *C. crista* and *C. pulcherrima*: The aerial parts of *C. crista* and *C. pulcherrima* were collected for Osmania University campus, Hyderabad in May, 2007 and were botanically identified. Voucher specimen (Nos. CC-150509 for *C. crista* and C.P-150509 for *C. pulcherrima*) are preserved in IIC herbarium.

The air dried plant material (2 kg) of *C. crista* was powdered and extracted thrice with $CHCl_3/MeOH$ (1:1) at room temperature. Each extraction was continued for 120 h using 5 L of solvent. The combined solvent was concentrated under reduced pressure to afford a thick brown gummy material (7 g). The residue was subjected to column chromatography over silica gel. The column was eluted with solvents of increasing polarity using

hexane and mixtures of hexane and EtOAc. The following compounds (amounts and eluting solvents are given) were obtained according to the increasing order of polarity: compound **2** (18 mg, hexane/EtOAc, 80:20), compound **1** (22 mg, hexane/EtOAc, 70:30), 7-methoxy bonducellin (32 mg, hexane/EtOAc, 60:40), 7-O-methyl sappanone (27 mg, hexane/EtOAc, 30:70) and sappanone (35 mg, hexane/EtOAc, 10:90).

The air dried and powdered plant material (2 kg) of *C. pulcherrima* was extracted thrice with CHCl₃/MeOH (1:1) at room temperature following the above method as described for extraction of *C. crista*. The concentrated extract (a thick brown gummy residue, 8 g) was subjected to column chromatography over silica gel. The column was eluted with solvents of increasing polarity using hexane and mixtures of hexane and EtOAc. The following compounds (amounts and eluting solvents are given) were isolated according to the increasing order of polarity: 2'-hydroxy-2,3,4',6'-tetramethoxy chalcone (9 mg, hexane/EtOAc, 80:20), isobonducellin (7 mg, hexane/EtOAc, 70:30), bonducellin (8 mg, hexane/EtOAc, 60:40), 5,7-dimethoxyflavone (10 mg, hexane/EtOAc, 50:50), 5,7-dimethoxy-3',4'-methylenedioxy flavone (12 mg, hexane/EtOAc, 40:60), and compound **3** (12 mg, EtOAc).

Compound **1** (6β-cinnamoyloxy-7β-acetoxycoumarin-5α-ol): White solid, mp 127–128 °C, $[\alpha]_D^{25} +63.8$ (c 0.3, CHCl₃), IR: 3448, 1721, 1636, 1456, 1238 cm⁻¹; ¹H and ¹³C NMR: Table 1; HREIMS: *m/z* 529.2578 [M+Na]⁺ (Calcd for C₃₁H₃₈O₆Na: *m/z* 529.2566).

Compound **2** (6β,7β-dibenzoyloxyycoumarin-5α-ol): White solid, mp 69–70 °C, $[\alpha]_D^{25} +13.7$ (c 0.3, CHCl₃); IR: 3446, 1724, 1459, 1280 cm⁻¹; ¹H and ¹³C NMR: Table 1; HREIMS: *m/z* 565.2555 [M+Na]⁺ (Calcd for C₃₄H₃₈O₆Na: *m/z* 565.2566).

Compound **3** (12-demethyl neocaesalpin F): White solid, mp 159–161 °C, $[\alpha]_D^{25} -56.6$ (c 1, CHCl₃); IR: 3425, 1722, 1449, 1276 cm⁻¹; ¹H and ¹³C NMR: Table 1; HREIMS: *m/z* 471.2395 [M+H]⁺ (Calcd for C₂₇H₃₅O₇: *m/z* 471.2382).

Hydrolysis of compounds **1** and **2**: Compounds **1** and **2** (10 mg each) were separately heated under reflux with 5% KOH in MeOH (2 mL) for 30 min. After usual work-up followed by purification of the reaction mixture over column chromatography both the compounds afforded **5** (5 mg from **1** and 6 mg from **2**) mp 106–107 °C, $[\alpha]_D^{25} +25.4$ (c 0.05, CHCl₃). The physical and spectral (IR, ¹H NMR and MS) properties of the compound were found to be identical to those reported in the literature.¹¹

11. Roach, J. S.; McLean, S.; Reynolds, W. F.; Trinto, W. F. *J. Nat. Prod.* **2003**, *68*, 1378.
12. Kinoshida, T.; Kaneko, M.; Noguchi, H.; Kitagawa, I. *Heterocycles* **1996**, *43*, 409.
13. Saitoh, T.; Sakashita, S.; Nakata, H.; Shimokawa, T.; Kinjo, J. E.; Yamahara, J.; Yamasaki, M.; Nohara, T. *Chem. Pharm. Bull.* **1986**, *34*, 2506.
14. Cabiddu, M. G.; Cabiddu, S.; Cadoni, E.; De Montij, S.; Fattuoni, C.; Melis, S.; Usai, M. *Synthesis* **2002**, 875.
15. Maheswara, M.; Siddaiah, V.; Rao, C. V. *Chem. Pharm. Bull.* **2006**, *54*, 1193.
16. Mosmann, T. *J. Immunol Methods* **1983**, *65*, 55.